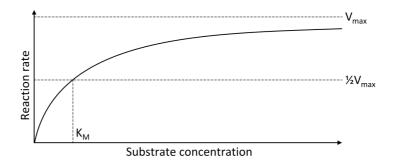
Enzyme- catalyzed reactions

The Michaelis-Menten equation describes the kinetics of enzyme-catalyzed reactions:

$$v = \frac{v_{max}[s]}{K_m + [s]}$$

Vmax represents the maximum rate achieved by the system, at saturating substrate concentration. The Michaelis constant Km is the substrate concentration at which the reaction rate is half of Vmax.



Gibbs free energy and equilibrium

By definition

 $\Delta G^0 = \Delta H^0 - T \Delta S^0$

At equilibrim

 $\Delta G^0 = \Delta H^0 - T \Delta S^0 = -RT lnK$

where ΔG^0 is the standard change in Gibbs free energy ΔH^0 is the standard change in enthalpy ΔS^0 is the standard change in entropy T is the temperature R is the gas constant

K is the equilibrium constant

Nearest Neighbor (NN) model for nucleic acids

This table presents the thermodynamic nearest neighbor (NN) parameters for Watson-Crick base pairs in 1 M NaCl.

TABLE 1 Nearest-neighbor thermodynamic parameters for DNA Watson-Crick pairs in 1 M NaCl^a

Propagation sequence	ΔH° (kcal mol ⁻¹)	ΔS° (e.u.)	ΔG_{37}° (kcal mol ⁻¹)
AA/TT	-7.6	-21.3	-1.00
AT/TA	-7.2	-20.4	-0.88
TA/AT	-7.2	-21.3	-0.58
CA/GT	-8.5	-22.7	-1.45
GT/CA	-8.4	-22.4	-1.44
CT/GA	-7.8	-21.0	-1.28
GA/CT	-8.2	-22.2	-1.30
CG/GC	-10.6	-27.2	-2.17
GC/CG	-9.8	-24.4	-2.24
GG/CC	-8.0	-19.9	-1.84
Initiation	+0.2	-5.7	+1.96
Terminal AT penalty	+2.2	+6.9	+0.05
Symmetry correction	0.0	-1.4	+0.43



The slash indicates the sequences are given in antiparallel orientation. (e.g., AC/TG means 5'-AC-3' is Watson-Crick base paired with 3'-TG-5'). The symmetry correction applies to only self-complementary duplexes. The terminal AT penalty is applied for each end of a duplex that has a terminal AT (a duplex with both end closed by AT pairs would have a penalty of +0.1 keal/mol for ΔG₂·2).

The following equation is used to predict the ΔG_T^0 at a different temperature, T:

$$\Delta G_T^0 = \Delta H^0 - T \Delta S^0$$

where T is in Kelvin, ΔH^0 is in cal/mol, and ΔS^0 is in units of $cal/K \cdot mol$ (entropy units, e.u.). ΔH^0 and ΔS^0 are assumed to be temperature independent; this is an excellent approximation for nucleic acids.

The NN Model.

The NN model for nucleic acids assumes that the stability of a given base pair depends on the identity and orientation of neighboring base pairs.

According to this model, the total $\Delta G_{Total(37)}^{0}$ is given by:

$$\Delta G_{Total(37)}^0 = \sum n_i \Delta G^0(i) + \Delta G^0(init /term \ G \star C) + \Delta G^0(init /term \ A \star T) + \Delta G^0(sym)$$

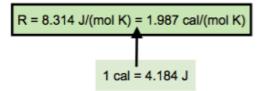
where $\Delta G^0(i)$ are the standard free-energy changes for the 10 possible Watson–Crick NNs (Table 1), n_i is the number of occurrences of each nearest neighbor (*i.e.* the number of pairs), i, and $\Delta G^0(sym)$ equals +0.43 kcal/mol (1cal =4.184J) if the duplex is self-complementary and zero if it is non-self-complementary.

Prediction of the Melting Temperature T_M.

 T_M is defined as the temperature at which half of the strands are in the double-helical state and half are in the "random-coil" state. The T_M is calculated from the predicted ΔH^0 and ΔS^0 , and the total oligonucleotide strand concentration C_T , by using the equation:

$$T_{\rm M} = \frac{\Delta H^0}{\Delta S^0 + R \ln C_{\rm T}}$$

where R is the gas constant (1.987 cal/K · mol).



To compute the T_M in Celsius degree:

$$T_M = \frac{\Delta H^0 \cdot 1000}{\Delta S^0 + R \ln C_T} - 273.15$$

since 0 K = -273.15 °C

where ΔH^0 is given in cal/mol.

Internal Single Mismatches

The nearest-neighbor model can be extended beyond the Watson-Crick pairs to include parameters for interactions between mismatches and neighboring base pairs. Table 2 provides the complete thermodynamic database for internal single mismatches.

TABLE 2 Nearest-neighbor ΔG_{32}° increments (kcal mol⁻¹) for internal single mismatches next to Watson-Crick pairs in 1 M NaCl*

Propagation sequence		Y			
	X	A	c	G	T
GX/CY	A	0.17	0.81	-0.25	WC
	C	0.47	0.79	WC	0.62
	G	-0.52	WC	-1.11	0.08
	T	WC	0.98	-0.59	0.45
CX/GY	A	0.43	0.75	0.03	WC
	C	0.79	0.70	WC	0.62
	G	0.11	WC	-0.11	-0.47
	T	WC	0.40	-0.32	-0.12
AX/TY	A	0.61	0.88	0.14	WC
	C	0.77	1.33	WC	0.64
	G	0.02	WC	-0.13	0.71
	T	WC	0.73	0.07	0.69
TX/AY	A	0.69	0.92	0.42	WC
	C	1.33	1.05	WC	0.97
	G	0.74	WC	0.44	0.43
	T	WC	0.75	0.34	0.68

 $[^]aWC$ indicates a Watson-Crick pair, which is given in Table 1. Error bars and ΔH^a and ΔS^a parameters are provided in the original references.

Antibody-antigens affinity and bond energy

If a monovalent antibody fragment is used for analysis, the equilibrium of antigen-antibody binding is defined as:

 $Antibody + Antigen \stackrel{K_a}{\Leftrightarrow} Complex$

where

$$K_a = \frac{[Complex]}{[Antibody][Antigen]}$$

Association and dissociation rate constants are defined as follows:

$$v_{ass} = k_{ass}[Antibody][Antigen]$$

 $v_{diss} = k_{diss}[Complex]$

where vass and vdiss represent the rates of association and dissociation, respectively, and kdiss represent the rate constants of association and dissociation, respectively. At equilibrium vass is equal to vdiss and the following equation is obtained:

$$K_a = \frac{k_{ass}}{k_{diss}}$$

 $K_a=rac{k_{ass}}{k_{diss}}$ The Gibbs' energy of formation (ΔG_0) of an antigen-antibody complex is given by:

$$\Delta G_0 = -RT \ln K_a$$

where R is the gas constant (1.987 cal/molK) and T is temperature.

The free energy of complex formation represents a balance between enthalpic (ΔH_0) and entropic (ΔS_0) forces as defined by the equation:

$$\Delta G_0 = \Delta H_0 - T \Delta S_0$$

In general, antigens and antibodies in solution have to overcome large entropic barriers before they can form a tight binding.